A mannan-conjugated myelin antigen inhibits the induction of CNS autoimmunity by CD11b⁺Ly6C^{hi} monocytes



Anastasia Dagkonaki ¹, Maria Avloniti ¹, Theodore Tselios ², Maria-Eleni Androutsou ³, Lesley Probert ^{1*} ¹ Laboratory of Molecular Genetics, Department of Immunology, Hellenic Pasteur Institute, Athens, Greece. ² Department of Chemistry, University of Patras, Rio, Patras, Greece.

³ Vianex SA, Varimbobi, Athens, Greece.

A

Introduction: Immature CD11b⁺Ly6Chi myeloid progenitors have been shown to be important for the development of experimental autoimmune encephalomyelitis (EAE). We recently showed that mannan-conjugated MOG (OM-MOG) ameliorates MOG-induced EAE by inducing a peripheral type 2 myeloid response. Here we investigate the involvement of Ly6Chi and PD-L1 in OM-MOG-induced immune tolerance.



Figure 1: CD11b⁺Ly6C^{hi} monocytes are critical for EAE onset and transiently depleted at disease peak. Gemcitabine depletes CD11b⁺Ly6C^{hi} and delays EAE onset. C57BL/6 mice were immunized with MOG (MOG35-55; 37µg) and treated with gemcitabine (GEM, i.p., 60mg/kg) at indicated timepoints (black arrows). (A) Percentages of CD11b+Ly6Chi cells in peripheral blood by FACS on days 0,8,11,13,15 and 28 dpi. (B) Mean clinical scores of EAE mice treated with GEM. Statistical analysis by Student's T test (A) and two-way ANOVA (B) is shown. (*p≤0.05, **p≤0.01, ***p≤0.001).



OM-MOG EAE /CD45

Figure 2: OM-MOG prevents spinal cord lesion formation by retaining Ly6Chi monocytes in the periphery.

C57BL/6 mice were immunized with MOG and treated with OM-MOG (i.d., 30 µg, upward arrows) on days 7,11 and 14, and with GEM (i.p., 60mg/kg) at indicated timepoints (downward arrows). (A) Mean clinical scores of mice treated with OM-MOG or OM-MOG+GEM at indicated time points. (B) Percentages of Ly6Chi in peripheral blood of the same mice as in A on days 0,8,11,13,15 and 28 post-immunization for EAE. (C) Whole mount spinal cord sections together with the surrounding bone taken at EAE peak (dpi18) and immunostained with antibodies against MBP and CD45. (D) Spinal cord sections from the same mice as in C immunostained with antibodies against laminin (LAM; red) and CD45 (green).

This research was co-financed by the European Union and Greek national funds through the Operational Program Competitiveness, Entrepreneurship and Innovation, under the call RESEARCH – CREATE – INNOVATE (project ΑΚΕΣΩ, project code:T1EDK-01859).





Figure 4: PD-L1 blockade reverses OM-MOG tolerance

C57BL/6 mice were immunized with MOG, untreated (grey squares) or treated with OM-MOG (i.d., 30 µg/injection) (black squares), and anti-PD-L1 antibody (i.p., 200µg/injection), or vehicle. (A) Mean clinical scores of mice treated with OM-MOG and/or anti-PD-L1 antibody at indicated time points. (B) Spinal cord sections taken from the day of sacrifice, stained with Luxol Fast Blue to show myelin (arrowheads show demyelinating lesions). Statistical analysis by Mann-Whitney test (*p≤0.05, EAE vs OM-MOG, # p≤0.05, OM-MOG vs PD-L1)

Conclusions

•Peripheral immature Ly6Chi monocytes are critical for EAE and their selective depletion by gemcitabine inhibits disease onset.

•OM-MOG induces immune tolerance by retaining Ly6Chi monocytes in the periphery, and is prolonged by gemcitabine.

•OM-MOG activates peripheral myeloid cells to produce PD-L1, which in turn is a critical mediator of OM-MOG immune tolerance.